

Glucose esters of caffeic acid from *Allium macrostemon* Bunge

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Abstract

Two new glucose esters of caffeic acid, 1-*O*-(*E*)-caffeoyl- β -D-glucopyranosyl-(1 \rightarrow 2)-[β -D-glucopyranosyl-(1 \rightarrow 6)]- β -D-glucopyranose (allimacronoid A-2, 1) and 1-*O*-(*E*)-caffeoyl- β -D-glucopyranosyl-(1 \rightarrow 4)-[β -D-glucopyranosyl-(1 \rightarrow 2)]-[β -D-glucopyranosyl-(1 \rightarrow 6)]- β -D-glucopyranose (allimacronoid B-2, 2) were isolated from the leaves of *Allium macrostemon* Bunge. The chemical structures were elucidated based on the analyses of the spectroscopic and chemical data.

Keywords : Amaryllidaceae, *Allium macrostemon*, caffeic acid, allimacronoid A-2, allimacronoid B-2

I Introduction

Allium macrostemon Bunge (Amaryllidaceae), known as wild onion, is widely distributed in East Asian countries. In Japan, the species is well known edible weed like other *Allium* species such as onion (*A. cepa* L.), garlic (*A. sativum* L.) and Welsh onion (*A. fistulosum* L.), etc. Some steroidal saponins have been isolated from the bulb¹⁾, which showed various biological activities²⁻⁵⁾ such as acute myocardial ischemia, hyperglycemia, hyperlipidemia and visceral obesity, etc. Nakane *et al.* reported the isolation and HPLC analysis of the flavonol glycoside constituents in the aerial part (leaf) of this plant⁶⁾. Recently, Usui *et al.* also isolated ferulic acid glucosides (1-*O*-(*E*)-feruloyl- β -D-glucopyranoside⁷⁾, 1-*O*-(*E*)-feruloyl- β -D-gentiobioside⁷⁾, tuberonoid A^{8, 9)}, allimacronoids A-D^{7, 9)}, 1-*O*-(*E*)-caffeoyl- β -D-sophoroside¹⁰⁾ and kaempferol glycosides¹⁰⁾ from the leaves of this species. In continuing systematic studies on the chemical constituents of *A. macrostemon*¹¹⁻¹³⁾, two new caffeic acid esters with oligo-glucose (1 and 2) were isolated from the leaves of the plant. The chemical structures of these compounds were elucidated from their spectroscopic and chemical data.

II Results and Discussion

A new compound 1 gave NH₄ adduct ion peak at *m/z* 684.2347 [M+NH₄]⁺, which corresponded to the molecular formula of C₂₇H₃₈O₁₉. The ¹H-NMR spectrum of 1 showed ABX-type (δ 6.78, *J*=8.0 Hz, 1H; δ 6.99 *J*=1.9, 8.0 Hz, 1H and δ 7.07, *J*=1.9 Hz, 1H) aromatic proton signals together with *trans*-olefinic (δ 6.31 and 7.65, *J*=15.8 Hz, each 1H) signals suggesting the presence of a caffeic acid moiety¹⁰⁾. Acid hydrolysis of 1 with 0.5 N HCl yielded *trans*-caffeic acid which was identified by HPLC analysis. The ¹H-NMR spectrum showed three sugar anomeric (δ 5.69, *J*=7.8 Hz, δ 4.58, *J*=7.8 Hz and δ 4.32, *J*=7.8 Hz) proton signals. In the ¹³C-NMR spectrum of 1 (Table 1), sugar signals (18 carbons) were also observed supporting the presence of three hexose moieties which were identified as D-glucose by TLC and HPLC analyses. These ¹H- and ¹³C-NMR data of 1 were almost identical to those of allimacronoid A⁹⁾ without only one methyl signal arising from ferulic acid unit of allimacronoid A. The characteristically downfield shifts of H-1 (δ 5.69) and H-6 (δ 4.16) signals observed in the ¹H-NMR spectrum (Table 1) suggested the pattern of the presence of the linkages between glucose C-1 - caffeic acid C-9, glucose C-2 - glucose C-1' and glucose C-6 - glucose C-1''^{7, 9, 10)}. These spectral data,